

## TRITERPENOID SAPONINS FROM *POLYSCIAS SCUTELLARIA*

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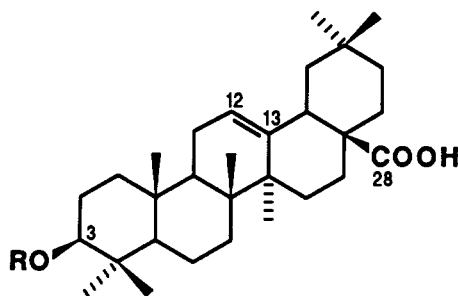
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**ABSTRACT.**—The leaves of *Polyscias scutellaria* afforded a new oleanolic glycoside, polysciasaponin P<sub>2</sub> [3], in addition to polysciasaponin P<sub>5</sub> [2], previously reported from *Panax zingiberensis*. The structures were established by chemical and spectroscopic means (fabms, <sup>13</sup>C nmr, gc-ms).

*Polyscias scutellaria* (Burm.f.) Fosb. (Araliaceae) is a bush used in the Pacific countries, especially in Vanuatu, as an anti-inflammatory (1). In previous work, we reported the isolation and structure determination of polysciasaponins P<sub>7</sub>, P<sub>6</sub>, and P<sub>4</sub> (2). In the present paper, we report the isolation and structure elucidation of polysciasaponins P<sub>5</sub> [2] and P<sub>2</sub> [3]. Compound 3 is a new triterpene glycoside. An EtOH extract of dried leaves was treated with several solvents (see Experimental); the *n*-BuOH fraction yielded a crude mixture of saponins. Separation of the *n*-BuOH extract by different chromatographic techniques afforded saponins P<sub>7</sub>, P<sub>6</sub>, P<sub>5</sub>, P<sub>4</sub>, and P<sub>2</sub>.

On acid hydrolysis polysciasaponins P<sub>5</sub> [2] and P<sub>2</sub> [3] afforded oleanolic acid [1] as the aglycone, identified by comparison with an authentic sample (tlc, ms, and gc-ms). The sugars obtained from the hydrolysates were identified by tlc and gc as glucuronic acid and glucose for 2 (1:1); glucuronic acid and glucose for 3 (1:2). Compounds 2 and 3 were not affected by alkaline treatment, indicating that they were monodesmosidic saponins. Mild acid hydrolysis of 2 gave 3-*O*-β-D-glucuronopyranosyl oleanolic acid, while mild acid hydrolysis of 3 gave 2 followed by 3-*O*-β-D-glucuronopyranosyl oleanolic acid.

Saponins 2 and 3 were submitted to fabms (negative ion mode) in order to establish the sugar sequence (3). The fabms spectrum of 2 showed a deprotonated molecular ion [M - H]<sup>-</sup> at *m/z* 793 (C<sub>42</sub>H<sub>65</sub>O<sub>14</sub>) and fragments at *m/z* 631 [M - H - 162]<sup>-</sup> and *m/z* 455 [M - H - 162 - 176]<sup>-</sup> corresponding to a subsequent loss of a glucosyl moiety and a glucuronic acid moiety, clearly indicating that the glucose was the terminal sugar. A deprotonated molecular ion [M - H]<sup>-</sup> at *m/z* 955 (C<sub>48</sub>H<sub>76</sub>O<sub>19</sub>) was character-



- 1 R=H
- 2 R=Glc(1→2) GlcA-
- 3 R=Glc(1→4) Glc(1→2) GlcA-  
GlcA = β-D-glucuronopyranosyl  
Glc = β-D-glucopyranosyl

ized in the fabms of **3**, corresponding to an oleanolic acid linked with two molecules of glucose and one of glucuronic acid. The fragments at  $m/z$  793 [ $M - H - 162$ ]<sup>-</sup> and  $m/z$  455 [ $M - H - 162 - 162 - 176$ ]<sup>-</sup> corresponding to a sequential loss of two glucosyl moieties and one glucuronic acid moiety, clearly indicated that the glucose molecules were the terminal sugars. The  $\beta$ -D-pyranosyl configurations of glucose and glucuronic acid in saponins **2** and **3** were derived from <sup>13</sup>C-nmr spectra (Table 1).

TABLE 1. <sup>13</sup>C-nmr Spectral Data of Polysciasaponin P<sub>2</sub> [**3**] in C<sub>5</sub>D<sub>5</sub>N ( $\delta$  ppm/TMS).

Carbon atom	
Oleanolic acid	
C-3 . . . . .	89
C-12 . . . . .	122.2
C-13 . . . . .	144.52
C-28 . . . . .	179.9
GlcA	
C-1' . . . . .	104.6
C-2' . . . . .	82.9
C-3' . . . . .	76.4
C-4' . . . . .	71.15
C-5' . . . . .	76.1
C-6' . . . . .	174.4
Glc	
C-1'' . . . . .	104.4
C-2'' . . . . .	74.5
C-3'' . . . . .	77.2
C-4'' . . . . .	80.5
C-5'' . . . . .	78.07
C-6'' . . . . .	62.2
Glc	
C-1''' . . . . .	104.3
C-1''' . . . . .	74.5
C-3''' . . . . .	77.25
C-4''' . . . . .	71.16
C-5''' . . . . .	77.85
C-6''' . . . . .	61.9

With Smith's degradation (periodate oxidation, NaBH<sub>4</sub> reduction, and partial hydrolysis), polysciasaponin P<sub>5</sub> [**2**] afforded oleanolic acid [**1**]. Thus, the molecule of glucose in **2** might be linked either with the 2'-OH or 4'-OH of glucuronic acid. In the <sup>13</sup>C-nmr spectrum of **2**, the chemical shifts at  $\delta$  82.9, 103.7, and 105.7 can be assigned to the C'-2 and the two anomeric carbon signals (C-1', C-1'') of  $\beta$ -(1 $\rightarrow$ 2) linked glucopyranose residues. The structure of polysciasaponin P<sub>5</sub> is established as 3-O- $\beta$ -[ $\beta$ -D-glucopyranosyl(1 $\rightarrow$ 2)- $\beta$ -D-glucuronopyranosyl] oleanolic acid.

With Smith's degradation, polysciasaponin P<sub>2</sub> [**3**] afforded oleanolic acid [**1**]. This result is not possible with a second glucose on the glucuronic acid. The <sup>13</sup>C-nmr sugar signals (Table 1) indicated that the  $\beta$ -D-glucopyranosyl is the terminal unit and that this sugar is attached at the position 4'' of the inner glucosyl. Thus, the structure of polysciasaponin P<sub>2</sub> is established as 3-O-[ $\beta$ -D-glucopyranosyl (1 $\rightarrow$ 4)- $\beta$ -D-glucopyranosyl (1 $\rightarrow$ 2)- $\beta$ -D-glucuronopyranosyl] oleanolic acid.

Polysciasaponin P<sub>2</sub> [**3**] is a new natural compound. Polysciasaponin P<sub>5</sub> [**2**] was ear-

lier reported from *Panax zingiberensis* (4) and *Panax japonicus* var. *angustifolius* (5); **2** has molluscicidal (6) and hemolytic activities (7).

## EXPERIMENTAL

**GENERAL EXPERIMENTAL PROCEDURES.**—Fabms were obtained on a VG Micromass ZAB-HF spectrometer in the negative ion mode; samples were suspended in polyethylene glycol.  $^{13}\text{C}$ -nmr spectra were recorded on a Bruker WP 200 in  $\text{C}_5\text{D}_5\text{N}$ . The chemical shifts are given in ppm. TMS was used as internal standard. Tlc was carried out on Si gel 60 (Merck). The tlc systems employed for saponins were *n*-BuOH–HOAc– $\text{H}_2\text{O}$  (4:1:1) (system 1), EtOAc–MeOH– $\text{H}_2\text{O}$  (60:15:6) (system 2), and  $\text{CHCl}_3$ –MeOH– $\text{H}_2\text{O}$  (60:30:4) (system 3). The systems employed for aglycones were  $\text{CHCl}_3$ –MeOH (97:3) (system 4) and  $\text{C}_6\text{H}_6$ –EtOAc (3:1) (system 5).

**PLANT MATERIAL.**—The leaves of *P. scutellaria* were collected from Vanuatu by P. Cabalion (ORSTOM). A voucher specimen is deposited in the Herbarium of Botany and Pharmacognosy, Faculté de Pharmacie, Lyon.

**EXTRACTION AND ISOLATION OF THE SAPONINS.**—Leaves (750 g) were extracted with hot EtOH. The EtOH extract was concentrated, diluted with  $\text{H}_2\text{O}$ , and extracted with  $\text{CHCl}_3$  to remove lipid material. The  $\text{H}_2\text{O}$  solution was extracted with EtOAc, then *n*-BuOH. The *n*-BuOH layer was evaporated to dryness to give a crude saponin fraction (21 g). The separation was carried out on a column packed with Si gel 60 (Merck) using a solvent gradient of  $\text{CHCl}_3$ –MeOH– $\text{H}_2\text{O}$  (70:2:2) to (70:40:10). Six fractions were collected. Fraction 2 gave polysciasaponin  $\text{P}_2$  [**3**] (80 mg); fraction 4 gave polysciasaponins  $\text{P}_3$  [**2**] and  $\text{P}_6$ . Saponins  $\text{P}_5$  and  $\text{P}_6$  were further separated on tlc (Si gel) with *n*-BuOH–HOAc– $\text{H}_2\text{O}$  (4:1:1) (detection with iodine) to give pure  $\text{P}_5$  (90 mg) and  $\text{P}_6$ .

**ACID HYDROLYSIS.**—The saponin [5 mg in MeOH– $\text{H}_2\text{O}$  (2:1) (5 ml)] was refluxed in 0.1 M HCl (5 ml) for 6 h. The aglycone was extracted with  $\text{CHCl}_3$  and identified with an authentic sample by tlc on Si gel and gc-ms after silylation. The aqueous layer was neutralized with *N,N*-octyl-methylamine and evaporated to dryness. The sugars were identified by gc after silylation and by tlc on Si gel using *n*-BuOH–*i*PrOH– $\text{H}_2\text{O}$  (100:60:20) and on cellulose using *n*-BuOH–pyridine–HCl (50:30:20); compounds were visualized by spraying with aniline hydrogen phthalate and then heating (110°, 10 min).

**PARTIAL ACID HYDROLYSIS.**—The saponin (5 mg) in MeOH (0.2 ml) and 1 M HCl (0.2 ml) was kept at 70°; after 80 min the mixture was separated by tlc on Si gel with *n*-BuOH–HOAc– $\text{H}_2\text{O}$  (4:1:1) to obtain the partially hydrolyzed products.

**ALKALINE HYDROLYSIS.**—Alkaline hydrolysis was performed at 80° for 3 h with 5 mg of saponin in 1 M NaOH (5 ml). After acidification by 1 M HCl (pH 5) monodesmoside was extracted with *n*-BuOH.

**SMITH'S DEGRADATION.**—To 5 mg of saponin was added 50 ml of  $\text{NaIO}_4$ , and the reaction mixture was allowed to stand for 48 h. The solution was treated with ethylene glycol (2 ml), then with  $\text{NaBH}_4$  (200 mg) for 15 h at 4°. The mixture was extracted with *n*-BuOH, then evaporated. The dry residue was treated with 0.5 M HCl for 7 h at 37°, then extracted with  $\text{CHCl}_3$  and analyzed by tlc on Si gel using *n*-BuOH–HOAc– $\text{H}_2\text{O}$  (4:1:1) and  $\text{CHCl}_3$ –MeOH (97:3).

**POLYSCIASAPONIN  $\text{P}_3$  [**2**].**—Tlc  $R_f$  0.46 (system 1), 0.21 (system 2), 0.12 (system 3); fabms  $m/z$   $[\text{M} - \text{H}]^-$  793,  $[\text{M} - \text{H} - 162]^-$  631,  $[\text{M} - \text{H} - 162 - 176]^-$  455;  $^{13}\text{C}$  nmr ( $\text{C}_5\text{D}_5\text{N}$ )  $\delta$  88.5 (C-3), 121.9 (C-12), 144.3 (C-13), 179.3 (C-28), 103.7 (C-1'), 105.7 (C-1''). Smith's degradation: **2** gave oleanolic acid [**1**].

**POLYSCIASAPONIN  $\text{P}_2$  [**3**].**—Tlc  $R_f$  0.31 (system 1), 0.09 (system 2), 0.05 (system 3); fabms  $m/z$   $[\text{M} - \text{H}]^-$  955,  $[\text{M} - \text{H} - 162]^-$  793,  $[\text{M} - \text{H} - 500]^-$  455;  $^{13}\text{C}$  nmr ( $\text{C}_5\text{D}_5\text{N}$ ) see Table 1. Smith's degradation: **3** gave oleanolic acid [**1**].

**ACID HYDROLYSIS.**—Saponins **2** and **3** gave oleanolic acid [**1**] identified by tlc  $R_f$  0.71 (system 4),  $R_f$  0.56 (system 5) and by gc-ms after silylation: ms  $m/z$   $[\text{M}]^+$  600,  $[\text{M} - \text{COOSi Me}]^+$  483,  $[\text{M} - 280]^+$  320,  $[\text{M} - 321]^+$  279,  $[\text{M} - 397]^+$  203,  $[\text{M} - 411]^+$  189. The sugars were identified by tlc and by gc after silylation. The sugar components were Glc–GlcA (1:1) for saponin **2** and Glc–GlcA (2:1) for saponin **3**.

**MILD ACID HYDROLYSIS.**—Compound **2** gave calenduloside E (3-*O*- $\beta$ -D-glucuronopyranosyl oleanolic acid) (8). Compound **3** gave **2**, then calenduloside E.

**ALKALINE HYDROLYSIS.**—Compounds **2** and **3** were not hydrolyzed.

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